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ORAL ABSTRACTS

631.MYELOPROLIFERATIVE SYNDROMES AND CHRONIC MYELOID LEUKEMIA: BASIC AND TRANSLATIONAL

Aberrant Activity of the Calcium Sensor STIM1 Underlies Congenital Platelet Disorders and Myeloproliferative Neoplasms

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This study was initiated following evaluation of a 32 year-old woman who presented with a history of thrombocytopenia identified in childhood who subsequently developed features of myelofibrosis (MF). A bone marrow biopsy demonstrated hypercellularity in conjunction with megakaryocyte hyperplasia and marked reticulin fibrosis. Molecular testing for *JAK2*, *CALR*, and *MPL* mutations was negative. Given the unusual association between congenital thrombocytopenia and MF in this patient, exome sequencing was performed, revealing a heterozygous R304W mutation in the coiled coil (CC) domain of *STIM1*. Activating mutations in the CC and EF hand domains of *STIM1* have been associated with Stormorken syndrome, a rare congenital platelet disorder associated with abnormal store operated calcium entry (SOCE). We subsequently identified a second patient with MF associated with a *STIM1* activating mutation. This individual was found to have severe thrombocytopenia at birth, and exome sequencing revealed a heterozygous S88G mutation in the EF hand of *STIM1*. A bone marrow biopsy obtained at 6 months of age revealed atypical megakaryocytes and grade 1-2 MF. A repeat biopsy at age 2 showed persistence of stable MF.

The unusual finding of MF in these two patients suggested the possibility of altered calcium signaling as a shared mechanism driving congenital platelet disorders such as Stormorken syndrome and myeloproliferative neoplasms (MPNs) including MF. In support of this notion, we identified elevated *STIM1* expression in MF vs normal megakaryocyte progenitors, as well as in platelets from patients with essential thrombocythemia (ET) vs healthy controls. Additionally, we found that *STIM1* expression was significantly elevated in CD34+ hematopoietic stem/progenitor cells (HSPCs) from both ET and MF patients vs healthy controls. Notably, *STIM1* expression was increased in both *JAK2* and *CALR*-mutant MF patients. Collectively, these findings provide evidence of aberrant *STIM1* expression in MPN patient cells.

To determine the functional role of *STIM1* in MPN disease development, colony assays and patient-derived xenograft (PDX) experiments were performed with MF patient CD34+ cells subjected to CRISPR ablation of *STIM1*. Strikingly discordant results were observed with *JAK2* vs *CALR*-mutant patient samples. Abrogation of *STIM1* in *CALR*-mutant CD34+ cells led to decreased colony formation, and NSGS mice engrafted in parallel with *STIM1*-targeted cells exhibited decreased human CD45+ cell engraftment in conjunction with prolonged survival. These findings suggest an important role for *STIM1* in *CALR*-mutant MPN disease phenotypes. In contrast, targeting of *STIM1* in *JAK2*-mutant CD34+ cells led to increased colony formation and exacerbated disease phenotypes in vivo as manifested by enhanced human CD45+ cell engraftment, worsened splenomegaly, and early lethality. Similar results were obtained in experiments utilizing pharmacologic inhibitors of SOCE

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activity. Taken together, these findings indicate that the consequences of aberrant STIM1 activity may be context-dependent relating to specific MPN driver mutations.

To expand these observations, we identified a separate cohort of 9 family members in Italy with Stormorken syndrome and confirmed *STIM1* EF hand mutations. In *ex vivo* megakaryocytic differentiation assays, cells from affected individuals exhibited a defect in proplatelet formation. These observations were corroborated by initial analyses of a newly generated *Stim1* R304W conditional knock-in mouse which recapitulated the characteristic thrombocytopenia found in patients with Stormorken syndrome.

In summary, this study represents the first demonstration of MF development in patients with Stormorken syndrome, thereby uncovering a previously unrecognized hallmark of altered calcium signaling via aberrant *STIM1* activation underlying Stormorken syndrome and MPNs. Our findings suggest distinct mechanisms relating to the interaction between *JAK2* vs *CALR* mutation and altered STIM1 activity. Further studies of these relationships may have important ramifications for potential therapeutic approaches targeting these pathways.

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